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*Graft* 2002; 5; 88

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# The Pharmacologic Treatment of Human Polyomavirus Infection

Julie Roskopf, William Fitzsimmons, Nasimul Ahsan, and David Laskow

Human polyomavirus (PV) infections are increasingly being recognized as a cause of significant morbidity and mortality in a subset of patients. Patients at highest risk for developing clinically significant disease due to polyomavirus include recipients of organ transplants and patients with human immunodeficiency virus type 1 (HIV-1) infection, due to impaired cellular immunity. BK virus (BKV) is most frequently associated with nephropathy and ureteral stenosis in renal transplant patients and nonhemorrhagic and hemorrhagic cystitis (HC) found predominantly in bone marrow transplant recipients. JC virus (JCV) is known to cause progressive multifocal leukoencephalopathy (PML) and is most commonly seen in patients who have HIV-1 infection. The treatment strategy for patients, regardless of the type of polyomavirus, is the judicious lowering of immunosuppression or the treatment of the underlying immunodeficiency disorder. In addition, patients may benefit from antiviral agents with activity against polyomaviruses. Cidofovir has been used successfully to treat patients with BKV-induced transplant nephropathy or BKV-induced HC. The nucleoside analogue vidarabine has also been used to treat BKV-induced HC. A different nucleoside analogue (cytarabine), interferons, highly active antiretroviral therapy (HAART), cidofovir, and the topoisomerase I inhibitors have all been used to treat AIDS-associated PML with variable success. Several other compounds, including retinoic acid and the malononitrilamide compound FK-778, may also prove to be useful antipolyomavirus compounds. It is clear that the optimal management of patients with polyomavirus infections is still unclear, but further research is ongoing and is likely to improve the care we are able to provide these patients.

## ABBREVIATIONS:

PV	Polyomavirus
HIV-1	Human immunodeficiency virus type 1
BKV	BK virus
HC	Hemorrhagic cystitis
JCV	JC virus
PML	Progressive multifocal leukoencephalopathy
HAART	Highly active antiretroviral therapy
DNA	Deoxyribonucleic acid
HSV	Herpes simplex virus
CMV	Cytomegalovirus
AIDS	Acquired immunodeficiency syndrome
IV	Intravenously
PCR	Polymerase chain reaction
PGE2	Prostaglandin E2
Ara-A	Adenine arabinoside or vidarabine
ACTG	AIDS Clinical Trials Group
SC	Subcutaneously
RNA	Ribonucleic acid

## Introduction

Polyomavirus (PV) is a nonenveloped, double-stranded deoxyribonucleic acid (DNA) virus that is a member of the *Polyomaviridae* family.<sup>1</sup> Human PV infections are due to BKV and JCV. Clinically significant disease due to PV is primarily found in patients with impaired cellular immunity such as recipients of organ transplants or patients with HIV-1 infection and is linked to the degree of overall immunosuppression.<sup>2,3</sup>

## BKV

BKV is most frequently associated with nephropathy (interstitial nephritis) and ureteral stenosis in renal transplant recipients, as well as nonhemorrhagic and hemorrhagic cystitis (HC), found predominantly in bone marrow transplant

recipients.<sup>3,4</sup> Clinical disease with BKV manifests principally in the genitourinary tract because the virus is known to remain latent in the kidney.<sup>3</sup> There is currently no standardized treatment available for patients diagnosed with BKV-associated disease. The most common treatment strategy is to drastically reduce or discontinue immunosuppressive drugs and treatments, if possible. This approach is designed to reduce viral replication but may be associated with risks such as acute rejection and graft loss in renal transplant patients, and patients must be monitored closely.<sup>5,6</sup>

## BKV-Induced Transplant Nephropathy

Certain risk factors, such as multiple acute rejection episodes and the availability of more potent immunosuppressive medications, have been associ-

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DOI: 10.1177/1522162802238461

ated with BKV nephropathy (typically, interstitial nephritis).<sup>7</sup> Patients often present with a rise in serum creatinine, necessitating further investigation. The diagnosis requires biopsy of the allograft and the use of either immunohistochemical analysis or in situ hybridization. These advanced techniques are needed to help distinguish if the tubular injury is due to the virus or the presence of concurrent acute rejection.<sup>6</sup>

A better understanding of how to manage BKV-induced nephropathy is slowly evolving. Historically, it was difficult to exclude concurrent acute rejection on biopsy; therefore, many patients were given intensified immunosuppressive regimens to treat acute rejection. This resulted in graft loss in a large number of patients, likely due to BKV (Table 1). Some authors have suggested that if subclinical acute rejection is found concomitantly on tissue biopsy, these patients may benefit from pulse steroid therapy followed by a reduction in baseline immunosuppression.<sup>2,14</sup> Although steroids may be of benefit, other authors have suggested it is best to avoid anti-T-cell agents such as OKT-3 and antithymocyte globulins after a diagnosis of BKV nephropathy has been made.<sup>7,13</sup>

Better diagnostic testing and more experience suggest that in the case of BKV nephropathy the mainstay of therapy is to lower the amount of maintenance immunosuppression so that the patient can overcome the viral infection.<sup>6,7,9,13</sup> Several approaches have included reducing or stopping azathioprine, mycophenolate mofetil, or sirolimus; using lower target concentrations of the calcineurin inhibitors (cyclosporine or tacrolimus); switching from one calcineurin inhibitor to another; or stopping the calcineurin inhibitor completely. This has resulted in a variety of different outcomes (Table 1).

Cidofovir (HPMPC: (S)-1-(3-hydroxy-2-phosphonylmethoxypropyl)cytosine) is an acyclic nucleoside phosphonate that is active against virtually all herpesviruses, as well as papovaviruses (polyomaviruses and papillomaviruses), adenoviruses, iridoviruses, and poxviruses (Fig. 1). Andrei and colleagues evaluated several different antiviral compounds, and cidofovir was found to be the most selective inhibitor of murine polyomavirus.<sup>20</sup> The spectrum of activity for cidofovir is very different from classic acyclic nucleoside analogues such as

### Key Points

- The development of clinically significant disease due to PV is linked to the degree of overall immunosuppression.
- The mainstay of therapy for treating BKV-induced transplant nephropathy is to lower the amount of maintenance immunosuppression so that the patient can overcome the viral infection.
- Cidofovir has been used successfully in the management of BKV nephropathy and may benefit a subset of patients who do not respond to a reduction in overall immunosuppression.
- BKV is also associated with HC in patients who have undergone either allogeneic bone marrow or stem cell transplant.
- JCV causes PML and is most commonly seen in patients with HIV-1 infection; several different antiviral agents have been used to try and treat PML, including Ara-A, interferons, HAART, and cidofovir.

acyclovir, penciclovir, and ganciclovir. In murine models, cidofovir is more effective than acyclovir against herpes simplex virus (HSV) infection and more effective than ganciclovir against cytomegalovirus (CMV) infection.<sup>21,22</sup> The active intracellular metabolite, cidofovir diphosphate, suppresses viral replication by the selective inhibition of viral DNA synthesis.<sup>23</sup> Cidofovir diphosphate accumulates in the cell (half-life = 65 h) and allows for a long-lasting antiviral effect with infrequent dosing. Cidofovir is currently approved for CMV retinitis in patients with acquired immunodeficiency syndrome (AIDS). It is often reserved for patients who are unresponsive to or have relapsed on intravenous ganciclovir or foscarnet.<sup>22,24</sup>

Recently, the successful use of cidofovir in 12 patients with BKV nephropathy has been described.<sup>25</sup> Cidofovir was administered as a single dose of 0.25 to 1 mg/kg every 2 to 3 weeks intravenously (IV) for a total of 1 to 4 doses. Low doses (5%–20% of the dose recommended for the treatment of CMV retinitis, given less often) were used because the

**Table 1 | OUTCOMES OF VARIOUS MANAGEMENT STRATEGIES IN PATIENTS WITH POSTTRANSPLANT BKV NEPHROPATHY**

SOURCE	DISEASE ONSET (MONTHS)	PATIENT NUMBER	PHARMACOLOGIC MANAGEMENT	OUTCOME/COMMENTS
Mathur et al. <sup>8</sup>	5-7	2	2 patients: Decrease IS	2/2 patients creatinine stabilized
Randhawa et al. <sup>5</sup>	Mean = 11.5 ± 13 Median = 9.1 Range = 1.5-66	22	2 patients: No intervention 12 patients: Given antirejection therapy 8 patients: Decrease IS	Diagnosed at nephrectomy 1/12 (8%) viral clearance 3/12 (25%) partial response 8/12 (67%) graft failure No graft loss Viral clearance in 3/6 (50%) patients
Howell et al. <sup>9</sup>	Mean = 9.3 ± 4.3 Range = 2.5-14.5	7	1 patient: Given antirejection therapy with increase IS 3 patients: Given antirejection therapy and later decrease IS 3 patients: Decrease IS	Diagnosed at nephrectomy 2/3 need dialysis soon 1/3 creatinine stabilized/improved Creatinine stabilized/improved
Nickeleit et al. <sup>2,10,11</sup> and Binet et al. <sup>12</sup>	Mean = 9.6 Range = 2.8-25	11	1 patient: Decrease IS 10 patients: Not clearly stated	Creatinine returned to baseline 5/10 creatinine increased but maintaining graft function 5/10 graft failure
Ahuja et al. <sup>13</sup>	Median = 9.5 Range = 6-32	10	10 patients: Given antirejection therapy and later decrease IS (concurrent AR on biopsy)	7/10 graft failure (6 rapid) 3/10 all functioning but follow-up < 1 year
Barri et al. <sup>7</sup>	Mean = 8 Range = 3-33	8	3 patients: Given antirejection therapy and decrease IS 4 patients: Decrease IS 1 patient: No intervention	2/3 creatinine stabilized 1/3 creatinine deteriorated 3/4 creatinine stabilized 1/4 graft failure Graft failure: Patient failed to follow up and stopped IS
Mayr et al. <sup>14</sup>	4	1	Given antirejection therapy and later decrease IS (concurrent AR on biopsy)	Creatinine stabilized
Hirsch et al. <sup>4</sup>	2.5	1	Decrease IS	Creatinine improved and virus clearance
Hussain et al. <sup>15</sup>	Not reported	14	7 patients: Given antilymphocyte therapy 7 patients: No intervention	7/7 graft failure 2/7 graft failure
Scantlebury et al. <sup>16</sup>	Mean = 13.4 Range = 2-32	34	12 patients: Given antirejection therapy and later decrease IS 9 patients: Decrease IS (concurrent AR on biopsy not treated) 13 patients: Decrease IS	3/12 (25%) creatinine stabilized 3/12 (25%) creatinine continues to rise 6/12 (50%) graft failure 4/9 (44%) creatinine stabilized 5/9 (56%) graft failure 6/13 (46%) creatinine improved 4/13 (31%) creatinine stabilized 2/13 (15%) graft failure 1/13 (8%) death with a functioning graft
Guardia et al. <sup>17</sup>	Not reported	8	8 patients: Decrease IS	3/6 had 1 or more AR episodes 4/6 creatinine improved 1/6 creatinine stabilized 1/6 moderate impairment of renal function 1/8 return to HD for recurrent glomerulonephritis 1/8 death with a functioning graft
Trofe et al. <sup>18</sup>	Kidney: Median = 19 Range = 7-40 Kidney/pancreas: Median = 24 Range = 8-28	10	10 patients: Decrease IS	1/10 developed mild steroid sensitive AR 6/10 creatinine increased ≥ 20% (with 1 graft failure) 4/10 creatinine stabilized 100% pancreas graft survival
Ramos et al. <sup>19</sup>	Mean = 12.8 Range = 2-52	67	52 patients: Decrease IS 15 patients: No intervention	8/52 (15.3%) developed AR 6/52 (11.5%) viral clearance 11/67 (16.4%) graft failure

IS, immunosuppression; AR, acute rejection; HD, hemodialysis.

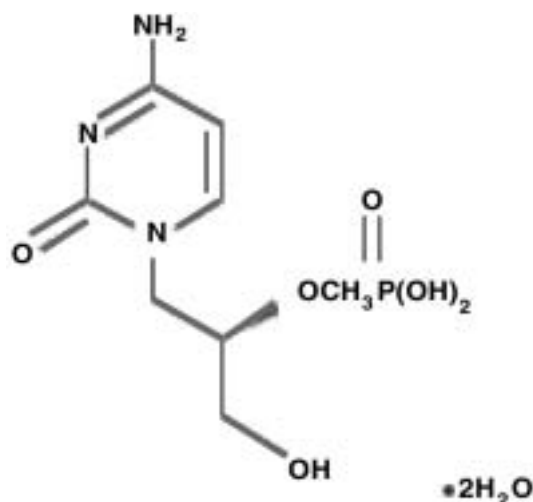


Figure 1. Chemical structure of cidofovir. Adapted from reference 23.

majority of cidofovir (75%–80%) is excreted unchanged in the kidney, and the goal is to treat virus localized in the kidney. Using quantitative polymerase chain reaction (PCR) for BKV, all patients showed clearance of viremia and either clearance or significant reduction of viruria. No lasting nephrotoxicity was seen with low-dose cidofovir, and the serum creatinine improved in all patients after treatment. Several patients have had recurrence of the viruria; therefore, after treatment, patients still need to be monitored closely. Vats<sup>25</sup> suggests that quantitative PCR testing can be used to diagnose and manage the course of BKV nephropathy and that cidofovir therapy may be beneficial in select patients, especially those who have not responded to a reduction in immunosuppression.

The most common clinical adverse events reported, when cidofovir is used to treat CMV retinitis, include nephrotoxicity (specifically proteinuria), nausea and vomiting, and fever.<sup>23</sup> Cidofovir has also been associated with neutropenia and intraocular inflammation (uveitis). To minimize the risk of nephrotoxicity when treating CMV retinitis, cidofovir is routinely given with high-dose oral probenecid (used to block the uptake of cidofovir by the proximal tubular cells) and saline prehydration (at least one liter of sodium chloride 0.9%) with each dose. When cidofovir is used to treat BKV nephropathy, however, lower doses of cido-

fovir are used (as mentioned above). This helps to minimize the risk for developing adverse events such as dose-dependent nephrotoxicity. Probenecid has not been used in patients being treated for BKV nephropathy, to allow for maximal excretion of the drug by the kidney, but prehydration with normal saline is still recommended. A second liter of fluid may be given over 1 to 3 hours during or after the cidofovir infusion if the patient will tolerate it.

Mayr and colleagues<sup>2,14</sup> have devised an algorithm for the screening, diagnosis, and monitoring of patients with BKV nephropathy. This algorithm includes the use of urine cytology to screen for decoy cells in high-risk patients. If there are > 5 decoy cells per 10 high-power fields found repeatedly, a plasma PCR to check for BKV is done. If this is repeatedly positive, then an allograft biopsy is done. If the diagnosis of BKV nephropathy is made, overall immunosuppression is decreased. If acute rejection is also present on the biopsy, the authors recommend the use of pulse steroids to treat the acute rejection, followed by a reduction in immunosuppression. We have devised an algorithm for the pharmacologic management of BKV nephropathy (Fig. 2). Many authors support the reduction or discontinuation of the antimetabolic agent in addition to reducing the target concentrations of the calcineurin inhibitor.<sup>7,9,13,16,18,19,26</sup> Cidofovir should be considered for patients who have persistent viremia/viruria despite a reduction in immunosuppression, especially if their renal function continues to worsen. Further research is needed to delineate if using this type of approach to treat BKV nephropathy is optimal to minimize progressive deterioration of renal function and ultimately prevent graft loss.

### BKV-Induced HC

HC is a well-defined complication following treatment with high-dose cyclophosphamide often used in bone marrow or stem cell transplant patients, and it typically occurs within 48 hours of infusion. HC associated with BKV differs because it most commonly occurs late and is long lasting. Despite the cause of HC, treatment is largely supportive and often includes hydration, alkalinization of the urine, bladder irrigation, pain management, antibiotics, and maintaining adequate platelet

Clinical disease with BKV manifests principally in the genitourinary tract because the virus is known to remain latent in the kidney.

Clinically significant disease due to PV is primarily found in patients with impaired cellular immunity, such as recipients of organ transplants or patients with HIV-1 infection, and is linked to the degree of overall immunosuppression.

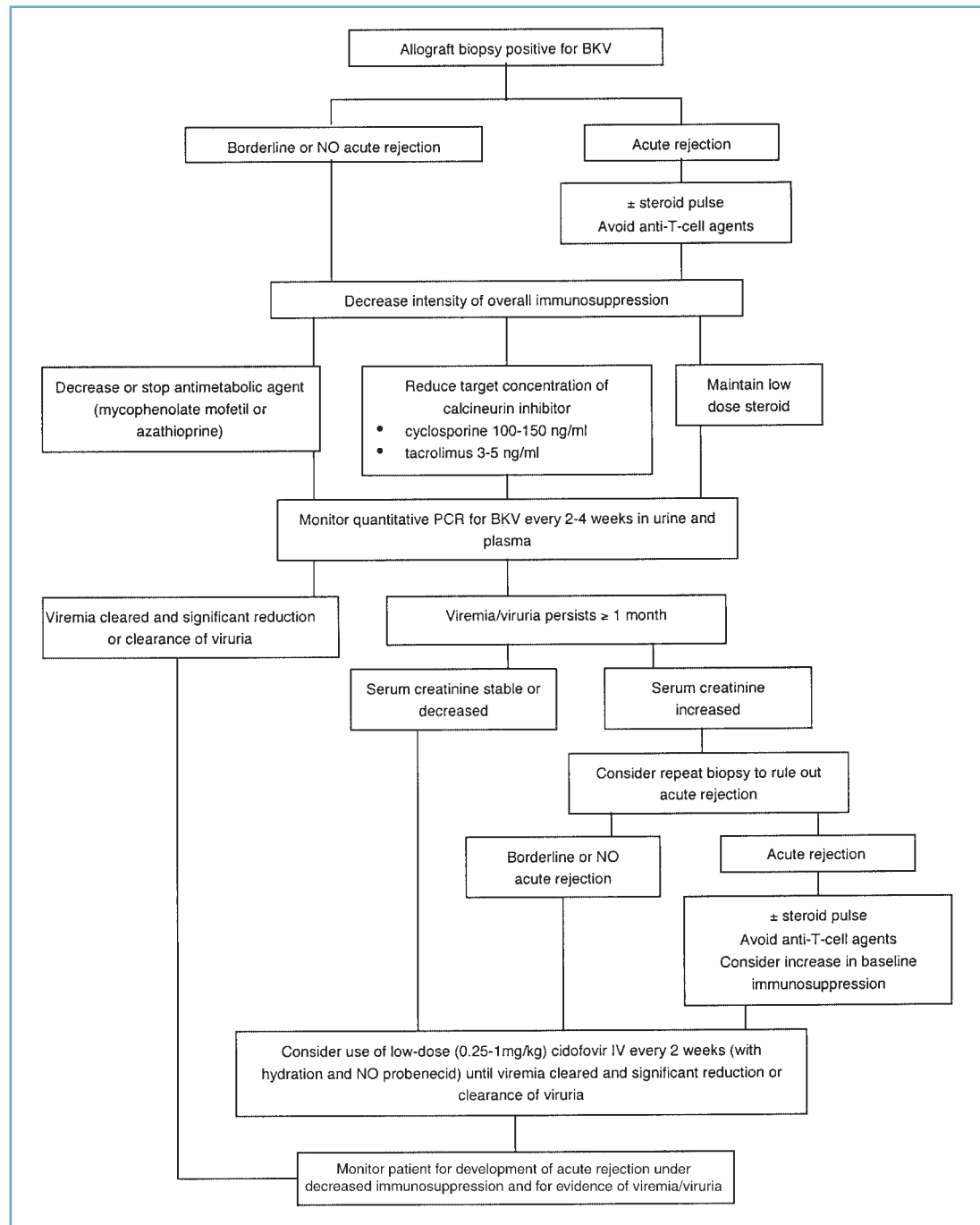


Figure 2. Algorithm for the pharmacologic management of patients with BKV nephropathy.

When cidofovir is used to treat BKV nephropathy, low doses of cidofovir are used to help minimize the risk for developing adverse events such as dose-dependent nephrotoxicity.

counts.<sup>3,27-29</sup> The intravesical installation of drugs such as formalin, aluminum, silver nitrate, and prostaglandin E2 (PGE2) may also be used for

more severe HC requiring more aggressive treatment, but bladder spasms tend to occur in most patients and may limit its usefulness.<sup>30,31</sup>

Several different drugs have been used to treat BKV-associated HC found in patients after either allogeneic bone marrow or stem cell transplant. The nucleoside analogue vidarabine (adenine arabinoside or Ara-A) is known to be active against several double-stranded DNA viruses.<sup>32</sup> It appears to exert its antiviral effects by interfering with the early steps in viral DNA synthesis.<sup>33</sup> Ara-A use has been described in several case reports. Chapman et al.<sup>34</sup> successfully treated a 23-year-old man with BKV-associated HC with Ara-A 10 mg/kg/day for 5 days, given IV over 12 h. Within 2 days, his symptoms improved, after 7 days he was symptom free, and within 20 days the virus was cleared from the urine. In another case series, Ara-A (10 mg/kg/day for 5 days, given IV over 2-3 h) was used in 1 patient with polyomavirus-associated cystitis and in 2 patients with asymptomatic polyoma viruria. With Ara-A therapy, the viral inclusion bodies in urinary sediments disappeared in all 3 patients. In 1 patient, the viruria recurred and was successfully cleared with another course of Ara-A.<sup>29</sup> In another case report, severe polyomavirus-associated HC was treated with Ara-A (10 mg/kg/day for 5 days, given intramuscularly), resulting in resolution of HC within 24 h of starting therapy and clearance of the virus from the urine after 4 days of treatment. Intramuscular Ara-A may be an alternative when patients have an adequate platelet count but do not have IV access.<sup>35</sup> The most common side effects of Ara-A include nausea, vomiting, diarrhea, increased liver function tests, headache, confusion, and tremor.<sup>33</sup> In the case reports above, Ara-A was associated with fatigue, nausea, vomiting, diarrhea, claustrophobia, and a transient increase in liver function tests.<sup>29,34,35</sup> Ara-A appears to be a safe and effective therapeutic alternative for BKV-associated HC, but parenteral Ara-A is no longer available in the United States. Ara-A was historically used to treat varicella-zoster and herpes simplex virus infections, but drugs such as acyclovir are now used to treat these types of viruses.<sup>33</sup>

There are several case reports describing the use of cidofovir for BKV-associated HC.<sup>27,28,36</sup> In 1 patient, after palliative treatment of HC failed, cidofovir was initiated at 5 mg/kg/week IV for 2 weeks, followed by 5 mg/kg IV every 2 weeks for a month (total 6 weeks) with concurrent probenecid to min-

imize the risk for nephrotoxicity. Symptoms improved after 2 weeks, and BK viruria cleared. The patient experienced nausea and vomiting thought to be secondary to probenecid and also a slight increase in serum creatinine that resolved in 1 week with IV hydration.<sup>27</sup> A different patient who had CMV reactivation in addition to BKV-associated HC was treated with cidofovir. Cidofovir is known to have potent activity against CMV in addition to polyomaviruses. Cidofovir was given as a single 5 mg/kg infusion and was repeated at 1 week and 3 weeks. Probenecid and IV hydration were also given simultaneously. As the BK viruria improved, the patient's symptoms of HC improved, and the CMV antigenemia also became negative. The serum creatinine became slightly elevated but returned to baseline.<sup>36</sup> In another patient, cidofovir was used for BKV-associated HC, but treatment failed and the patient underwent cystectomy.<sup>28</sup> It appears that cidofovir is safe and effective in this setting of BKV and should be considered for patients who fail conventional methods for controlling HC.

### JCV and PML

JCV causes progressive multifocal leukoencephalopathy (PML), a demyelinating disease of the central nervous system. PML is most commonly seen in patients who have HIV-1 infection, with the average survival after diagnosis ranging from approximately 2.5 to 4 months.<sup>37</sup> Cases of PML have also been described in patients with lymphoproliferative disorders, inherited primary immunodeficiency diseases, and exposure to high-dose chemotherapy, as well as in patients on prolonged immunosuppressive therapy (i.e., renal, liver, and heart allograft recipients and patients with rheumatoid arthritis).<sup>38-40</sup>

Currently, no effective therapy for PML has been clearly established, but progress has been made over the years due to a better understanding of the disease. The general treatment strategy is to minimize or discontinue immunosuppression, as described earlier in patients with BKV, or to try and correct the underlying immunodeficiency.

Historically, different drugs have been used to try and treat PML. The antiviral agent Ara-A has been used in an attempt to block JCV DNA replication.

PML is a demyelinating disease due to JCV, seen in immunosuppressed hosts.

In a case report by Rand et al.,<sup>32</sup> Ara-A did not appear to provide any significant clinical benefit when used systemically for 2 patients with advanced PML. A different nucleoside analogue, cytarabine (cytosine arabinoside or Ara-C), has been used frequently to treat PML. The efficacy of Ara-C has been supported by some uncontrolled studies but not by others.<sup>41-45</sup> Based on in vitro data showing that Ara-C could block JCV DNA replication without overt toxicity to glial cells, Ara-C was studied more formally.<sup>46</sup> The AIDS Clinical Trials Group (ACTG) Study 243 was a multicenter, open-label trial that was conducted to evaluate the use of Ara-C in HIV-infected patients with biopsy-confirmed PML.<sup>37</sup> The study was designed to enroll 90 patients, but only 64 patients were enrolled, and 57 patients were evaluated. Patients were randomly assigned to one of three treatment groups, each lasting 24 weeks (Fig. 3).

Twenty-two patients (39%) died during the study, and only 7 patients (12%) completed the 24-week study. The study closed early after 24 months, after finding that there was no statistical difference in the survival rates between the three treatment groups ( $p = 0.85$ ). The major toxicity associated with Ara-C therapy was hematologic (lower hemoglobin and platelet counts), and it occurred more often in the intravenous Ara-C group. More recently, Levy et al.<sup>47</sup> have suggested that Ara-C failure in the ACTG Trial 243 was due to inadequate drug delivery to target cells in the brain. They have proposed that alternative methods used to infuse Ara-C directly into the brain may prove efficacious.

Early reports also describe the use of interferons that possess antiviral and immunomodulating activity. Tashiro and coauthors<sup>48</sup> reported the use of intrathecal beta-interferon therapy in a single patient that resulted in stabilization of her neurological status. More recently, Huang and colleagues<sup>49</sup> reported results of an open-label study in HIV-associated PML patients. Untreated patients ( $n = 32$ ) were compared to patients ( $n = 21$ ) who received alpha-interferon (3 weeks minimum of either 3 million units subcutaneously [SQ] every day or 5 million units SQ three times per week). The median survival time was significantly prolonged in treated patients (325 days) versus untreated patients (121 days) ( $p < 0.001$ ). Side effects in the treatment

HAART has emerged as an essential component of treatment for patients with PML in an effort to restore the patient's inherent ability to mount an immune response against JCV.

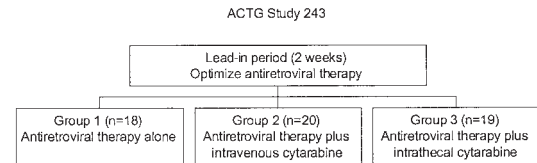


Figure 3. Evaluating the use of cytarabine in HIV-infected patients with progressive multifocal leukoencephalopathy (PML). Adapted from reference 37.

group occurred in 19% of patients and included leukopenia, pancytopenia, depression, and fatigue. The authors concluded that alpha-interferon should be studied more formally in a randomized clinical trial.

Improved survival has been reported for PML patients who receive highly active antiretroviral therapy (HAART).<sup>50,51</sup> This therapy is designed to achieve an undetectable HIV ribonucleic acid (RNA) viral load and an increase in CD4 cell count and often correlates with a patient's inherent ability to mount an immune response. HAART has emerged as an essential component of treatment for patients with PML; however, reports of PML exist in patients who are maximized on HAART therapy, and worsening of PML (both clinically and radiologically) has been reported in patients upon the initiation of HAART, suggesting an inflammatory reaction had occurred.<sup>52,53</sup> In fact, despite HAART, the incidence of PML has been relatively constant.<sup>54</sup> This has underscored the importance of identifying antiviral medications that are effective against JCV because reversing immunosuppression is not enough to alter the morbidity and mortality associated with PML in all patients.<sup>55</sup>

There have been numerous case reports describing the use of cidofovir for PML.<sup>24,56-63</sup> A multicenter observational study was conducted to assess if HAART plus cidofovir therapy was more effective than HAART alone in patients with AIDS-related PML.<sup>64</sup> Group A consisted of 27 patients treated with HAART alone, and group B consisted of 16 patients treated with HAART plus cidofovir. The probability of survival at 1 year was 0.29 in the HAART-alone group and 0.61 in the HAART-plus-cidofovir group ( $p = 0.02$ ). Evaluation at 2 months revealed that 5 of 12 patients tested (42%) in group A had undetectable JCV DNA in the cerebrospinal fluid (CSF), compared to 7 of 8 patients



(87%) in group B ( $p = 0.04$ ). In group B, there was a single patient who developed significant proteinuria that resolved after discontinuation of cidofovir. One other observational study also found that the use of cidofovir was significantly associated with survival, and another found that cidofovir was not associated with improved survival.<sup>65,66</sup> It is clear that further research is needed to develop the optimal strategy for treating patients with PML, but cidofovir appears to be a very reasonable adjunctive treatment option at this time. It may be especially useful in patients who have no improvement or progression of symptoms associated with PML while on HAART or in patients who cannot tolerate HAART.

Other possible therapies for PML include the use of topoisomerase I inhibitors, such as ironotecan, camptothecin, and topotecan. In vitro data indicate that camptothecin and topotecan can effectively block DNA replication of JCV in glial cells.<sup>67</sup> Topotecan and ironotecan are both synthetic analogues of camptothecin that have been clinically more useful with less severe side effects. Both of these drugs have been used successfully to treat AIDS-associated PML.<sup>68-70</sup> These types of drugs may be considered for the treatment of PML in both non-AIDS and AIDS patients, but optimal dosing and duration of therapy need to be investigated further.

### Conclusion

It is clear that the development of clinically significant disease with polyomavirus is related to the degree of overall immunosuppression. The key to improving clinical outcomes is the prompt diagnosis and treatment of patients with polyomavirus. In addition to supportive care, the mainstay of therapy is currently aimed at improving immunocompetence. Specific compounds, including cidofovir, the nucleoside analogues Ara-A and Ara-C, and the topoisomerase I inhibitors such as topotecan, have been used clinically to treat patients with BK or JC virus. Other agents, including retinoic acid, DNA gyrase inhibitors, 5'-bromo-2'-deoxyuridine, and the malononitrilamide compound FK-778, have shown antipolyomavirus activity in vitro but have not been tested in clinical trials.<sup>5,71,72</sup> In the future, these compounds may add to the current arma-

mentarium of agents used to treat polyomaviruses, and further research will aid in defining the optimal management for patients requiring treatment for clinically significant polyomavirus infections.

### REFERENCES

1. Major EO. Human polyomavirus. In: Knipe DM, Howley PM, Griffin DE, et al., editors. *Fields virology*. 4th ed. Philadelphia: Lippincott Williams and Wilkins; 2001. p. 2175-96.
2. Nicleleit V, Hirsch HH, Zeiler M, et al. BK-virus nephropathy in renal transplants-tubular necrosis, MHC-class II expression and rejection in a puzzling game. *Nephrol Dial Transplant* 2000;15(3):324-32.
3. Reploeg MD, Storch GA, Clifford DB. BK virus: a clinical review. *Clin Infect Dis* 2001;33(2):191-202.
4. Hirsch HH, Mohaupt M, Klimkait T. Prospective monitoring of BK virus load after discontinuing sirolimus treatment in a renal transplant patient with BK virus nephropathy. *J Infect Dis* 2001;184(11):1494-6.
5. Mylonakis E, Goes N, Rubin RH, et al. BK virus in solid organ transplant recipients: an emerging syndrome. *Transplantation* 2001;72(10):1587-92.
6. Randhawa PS, Finkelstein S, Scantlebury V, et al. Human polyoma virus-associated interstitial nephritis in the allograft kidney. *Transplantation* 1999;67(1):103-9.
7. Barri YM, Ahmad I, Ketel BL, et al. Polyoma viral infection in renal transplantation: the role of immunosuppressive therapy. *Clin Transplant* 2001;15(4):240-6.
8. Mathur VS, Olson JL, Darragh TM, et al. Polyomavirus-induced interstitial nephritis in two renal transplant recipients: case reports and review of the literature. *Am J Kidney Dis* 1997;29(5):754-8.
9. Howell DN, Smith SR, Butterly DW, et al. Diagnosis and management of BK polyomavirus interstitial nephritis in renal transplant recipients. *Transplantation* 1999;68(9):1279-88.
10. Nicleleit V, Klimkait T, Binet IF, et al. Testing for polyomavirus type BK DNA in plasma to identify renal-allograft recipients with viral nephropathy. *N Engl J Med* 2000;342(18):1309-15.
11. Nicleleit V, Hirsch HH, Binet IF, et al. Polyomavirus infection of renal allograft recipients: from latent infection to manifest disease. *J Am Soc Nephrol* 1999;10(5):1080-9.
12. Binet I, Nicleleit V, Hirsch HH, et al. Polyomavirus disease under new immunosuppressive drugs: a cause of renal graft dysfunction and graft loss. *Transplantation* 1999;67(6):918-22.
13. Ahuja M, Cohen EP, Dayer AM, et al. Polyoma virus infection after renal transplantation: use of immunostaining as a guide to diagnosis. *Transplantation* 2001;71(7):896-9.
14. Mayr M, Nicleleit V, Hirsch HH, et al. Polyomavirus BK nephropathy in a kidney transplant recipient: critical issues of diagnosis and management [computer file]. *Am J Kidney Dis* 2001;38(3):E13.
15. Hussain S, Bresnahan B, Cohen E, et al. Rapid kidney allograft failure is associated with antilymphocyte therapy in patients with polyoma virus interstitial nephritis [abstract]. *Am J Transplantation* 2001;1(suppl. 1):271.
16. Scantlebury V, Shapiro R, Justin G, et al. Graft function after diagnosis of BK virus in adult kidney transplant recipients under tacrolimus-based immunosuppression [abstract]. *Am J Transplantation* 2001;1(suppl. 1):404.
17. Guardia O, Re L, Rial M, et al. Long term clinical evolution of patients with BK virus infection after renal transplantation [abstract]. *J Am Soc Nephrol* 2001;12:932A.
18. Trofe J, Cavallo T, First M, et al. Polyomavirus in kidney and kidney pancreas transplantation: minimizing renal allograft injury/loss by defined immunosuppression reduction and histologic monitoring [abstract]. *Am J Transplantation* 2002;2(suppl. 3):292.
19. Ramos E, Weir M, Schweitzer E, et al. Clinical course of patients with polyoma virus nephropathy (PVN): a single center experience [abstract]. *Am J Transplantation* 2002;2(suppl. 3):294.

20. Andrei G, Snoeck R, Vandeputte M, et al. Activities of various compounds against murine and primate polyomaviruses. *Antimicrob Agents Chemother* 1997;41(3):587-93.
21. De Clercq E. Acyclic nucleoside phosphonates in the chemotherapy of DNA virus and retrovirus infections. *Intervirology* 1997;40(5-6):295-303.
22. De Clercq E. In search of a selective antiviral chemotherapy. *Clin Microbiol Rev* 1997;10(4):674-93.
23. Package insert. Vistide® (cidofovir injection), Foster City, CA; Gilead Sciences, Inc., September 2000. Available from: <http://www.gilead.com/wt/sec/vistide>.
24. Portilla J, Boix V, Roman F, et al. Progressive multifocal leukoencephalopathy treated with cidofovir in HIV-infected patients receiving highly active anti-retroviral therapy. *J Infect* 2000;41(2):182-4.
25. Vats A. Viral load monitoring in BK virus-associated nephropathy. *Transplant News & Issues* 2002;3(1):S17-9.
26. Loertscher R, Suri R, Lipman M, et al. Deliberate reduction of immunosuppression benefits patients with polyoma BK virus infection in kidney allografts [abstract]. *Am J Transplantation* 2002;2(suppl. 3):262.
27. Gonzalez-Fraile MI, Canizo C, Caballero D, et al. Cidofovir treatment of human polyomavirus-associated acute haemorrhagic cystitis. *Transplant Infect Dis* 2001;3(1):44-6.
28. Garderet L, Bittencourt H, Sebe P, et al. Cystectomy for severe hemorrhagic cystitis in allogeneic stem cell transplant recipients. *Transplantation* 2000;70(12):1807-11.
29. Kawakami M, Ueda S, Maeda T, et al. Vidarabine therapy for virus-associated cystitis after allogeneic bone marrow transplantation. *Bone Marrow Transplant* 1997;20(6):485-90.
30. Goddard AG, Saha V. Late-onset hemorrhagic cystitis following bone marrow transplantation: a case report. *Pediatr Hematol Oncol* 1997;14(3):273-5.
31. Laszlo D, Bosi A, Guidi S, et al. Prostaglandin E2 bladder instillation for the treatment of hemorrhagic cystitis after allogeneic bone marrow transplantation. *Haematologica* 1995;80(5):421-5.
32. Rand KH, Johnson KP, Rubinstein LJ, et al. Adenine arabinoside in the treatment of progressive multifocal leukoencephalopathy: use of virus-containing cells in the urine to assess response to therapy. *Ann Neurol* 1977;1(5):458-62.
33. Hoffmann R, Lewis I. Vidarabine (drug evaluation). In: Hutchinson TA, Shahan DR, editors. DRUGDEX® system. Greenwood Village, CO: MICROMEDEX (edition expires 9/2002).
34. Chapman C, Flower AJ, Durrant ST. The use of vidarabine in the treatment of human polyomavirus associated acute haemorrhagic cystitis. *Bone Marrow Transplant* 1991;7(6):481-3.
35. Seabra C, Perez-Simon JA, Sierra M, et al. Intra-muscular vidarabine therapy for polyomavirus-associated hemorrhagic cystitis following allogeneic hemopoietic stem cell transplantation. *Bone Marrow Transplant* 2000;26(11):1229-30.
36. Held TK, Biel SS, Nitsche A, et al. Treatment of BK virus-associated hemorrhagic cystitis and simultaneous CMV reactivation with cidofovir. *Bone Marrow Transplant* 2000;26(3):347-50.
37. Hall CD, Dafni U, Simpson D, et al. Failure of cytarabine in progressive multifocal leukoencephalopathy associated with human immunodeficiency virus infection. AIDS Clinical Trials Group 243 Team. *N Engl J Med* 1998;338(19):1345-51.
38. Aksamit A. Central nervous system. In: Bowden R, Ljungman P, Paya CV, editors. *Transplant infections*. Philadelphia: Lippincott-Raven; 1998. p. 133-52.
39. Shah K. Polyomaviruses. In: Fields BN, Knipe DM, Knipe PM, et al., editors. *Fields virology*. 3rd ed. Philadelphia: Lippincott-Raven; 1996. p. 2027-43.
40. Re D, Bamborschke S, Feiden W, et al. Progressive multifocal leukoencephalopathy after autologous bone marrow transplantation and alpha-interferon immunotherapy. *Bone Marrow Transplant* 1999;23(3):295-8.
41. Antinori A, De Luca A, Ammassari A, et al. Failure of cytarabine and increased JC virus-DNA burden in the cerebrospinal fluid of patients with AIDS-related progressive multifocal leukoencephalopathy. *AIDS* 1994;8(7):1022-4.
42. de Truchis P, Flament-Saillour M, Urtizberea JA, et al. Inefficacy of cytarabine in progressive multifocal leukoencephalopathy in AIDS. *Lancet* 1993;342(8871):622-3.
43. Guarino M, D'Alessandro R, Rinaldi R, et al. Progressive multifocal leukoencephalopathy in AIDS: treatment with cytosine arabinoside. *AIDS* 1995;9(7):819-20.
44. Portegies P, Algra PR, Hollak CE, et al. Response to cytarabine in progressive multifocal leukoencephalopathy in AIDS. *Lancet* 1991;337(8742):680-1.
45. Nicoli F, Chave B, Peragut JC, et al. Efficacy of cytarabine in progressive multifocal leukoencephalopathy in AIDS. *Lancet* 1992;339(8788):306.
46. Hou J, Major EO. The efficacy of nucleoside analogs against JC virus multiplication in a persistently infected human fetal brain cell line. *J Neurovirol* 1998;4(4):451-6.
47. Levy RM, Major E, Ali MJ, et al. Convection-enhanced intraparenchymal delivery (CEID) of cytosine arabinoside (AraC) for the treatment of HIV-related progressive multifocal leukoencephalopathy (PML). *J Neurovirol* 2001;7(4):382-5.
48. Tashiro K, Doi S, Moriwaka F, et al. Progressive multifocal leukoencephalopathy with magnetic resonance imaging verification and therapeutic trials with interferon. *J Neurol* 1987;234(6):427-9.
49. Huang SS, Skolasky RL, Dal Pan GJ, et al. Survival prolongation in HIV-associated progressive multifocal leukoencephalopathy treated with alpha-interferon: an observational study. *J Neurovirol* 1998;4(3):324-32.
50. Skiest DJ. Focal neurological disease in patients with acquired immunodeficiency syndrome. *Clin Infect Dis* 2002;34(1):103-15.
51. Clifford DB, Yiannoutsos C, Glicksman M, et al. HAART improves prognosis in HIV-associated progressive multifocal leukoencephalopathy. *Neurology* 1999;52(3):623-5.
52. Weiner SM, Laubenberger J, Muller K, et al. Fatal course of HIV-associated progressive multifocal leukoencephalopathy despite successful highly active antiretroviral therapy. *J Infect* 2000;40(1):100-2.
53. Miralles P, Berenguer J, Lacruz C, et al. Inflammatory reactions in progressive multifocal leukoencephalopathy after highly active antiretroviral therapy. *AIDS* 2001;15(14):1900-2.
54. Sacktor N, Lyles RH, Skolasky R, et al. HIV-associated neurologic disease incidence changes: Multicenter AIDS Cohort Study, 1990-1998. *Neurology* 2001;56(2):257-60.
55. Dworkin MS, Wan PC, Hanson DL, et al. Progressive multifocal leukoencephalopathy: improved survival of human immunodeficiency virus-infected patients in the protease inhibitor era. *J Infect Dis* 1999;180(3):621-5.
56. Blick G, Whiteside M, Grieger P, et al. Successful resolution of progressive multifocal leukoencephalopathy after combination therapy with cidofovir and cytosine arabinoside. *Clin Infect Dis* 1998;26(1):191-2.
57. Sadler M, Chinn R, Healy J, et al. New treatments for progressive multifocal leukoencephalopathy in HIV-1-infected patients. *AIDS* 1998;12(5):533-5.
58. Brambilla AM, Castagna A, Novati R, et al. Remission of AIDS-associated progressive multifocal leukoencephalopathy after cidofovir therapy. *J Neurol* 1999;246(8):723-5.
59. Martinez AC, Lopez AG, Garcia IG. Successful resolution of progressive multifocal leukoencephalopathy after combination therapy with cidofovir and cytosine arabinoside. *Clin Infect Dis* 2000;30(1):234.
60. De Luca A, Fantoni M, Tartaglione T, et al. Response to cidofovir after failure of antiretroviral therapy alone in AIDS-associated progressive multifocal leukoencephalopathy. *Neurology* 1999;52(4):891-2.
61. Cardenas RL, Cheng KH, Sack K. The effects of cidofovir on progressive multifocal leukoencephalopathy: an MRI case study. *Neuroradiology* 2001;43(5):379-82.
62. Herrero-Romero M, Cordero E, Lopez-Cortes LF, et al. Cidofovir added to highly active antiretroviral therapy in AIDS-associated progressive multifocal leukoencephalopathy. *AIDS* 2001;15(6):809.
63. Razonable RR, Aksamit AJ, Wright AJ, et al. Cidofovir treatment of progressive multifocal leukoencephalopathy in a patient receiving highly active antiretroviral therapy. *Mayo Clin Proc* 2001;76(11):1171-5.

64. De Luca A, Giancola ML, Ammassari A, et al. Potent anti-retroviral therapy with or without cidofovir for AIDS-associated progressive multifocal leukoencephalopathy: extended follow-up of an observational study. **J Neurovirol** 2001;7(4):364-8.
65. Berenguer J, Miralles P, Arrizabalaga J, et al. Clinical course and prognostic factors of AIDS-associated progressive multifocal leukoencephalopathy (PML) in patients treated with HAART [abstract 10]. Paper presented at the 8th Conference on Retroviruses and Opportunistic Infections, Chicago, February 2001.
66. Marra C, Rajcic N, Barker DE, et al. Prospective pilot study of cidofovir for HIV-associated progressive multifocal leukoencephalopathy (PML) [abstract 596]. Paper presented at the 8th Conference on Retroviruses and Opportunistic Infections, Chicago, February 2001.
67. Kerr DA, Chang CF, Gordon J, et al. Inhibition of human neurotropic virus (JCV) DNA replication in glial cells by camptothecin. **Virology** 1993;196(2):612-8.
68. Dupont B, Fish D, McGuire D, et al. 21-day continuous infusion of topotecan in AIDS-associated progressive multifocal leukoencephalopathy (PML) [abstract 597]. Paper presented at the 8th Conference on Retroviruses and Opportunistic Infections, Chicago, February 2001.
69. O'Reilly S. Efficacy of camptothecin in progressive multifocal leukoencephalopathy. **Lancet** 1997;350(9073):291.
70. Vollmer-Haase J, Young P, Ringelstein EB. Efficacy of camptothecin in progressive multifocal leukoencephalopathy. **Lancet** 1997;349(9062):1366.
71. Chen Y, Freund R, Listerud M, et al. Retinoic acid inhibits transformation by preventing phosphatidylinositol 3-kinase dependent activation of the c-fos promoter. **Oncogene** 1999;18(1):139-48.
72. Snoeck R, Andrei G, Lijja H, et al. Activity of malononitrilamide compounds against murine and simian polyomavirus [abstract]. Paper presented at the 5th International Conference on New Trends in Clinical and Experimental Immunosuppression, Geneva, Switzerland, February 2002. Available from: <http://www.kenes.com/immuno/abstracts/67.doc>.