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Expression and Function of CD40 on Hematopoietic and Non-Hematopoietic Cells

Cees van Kooten, Simone de Haij, Leendert C. Paul and Mohamed R. Daha

CD40 is a cell surface receptor which belongs to the TNF-R family. It was first identified and functionally characterized on B lymphocytes. The ligand for CD40 (CD40-L/CD154) is expressed mainly on activated T cells and plays a pivotal role in T cell-dependent B cell responses. However, in recent years it has become clear that CD40 has a much broader expression pattern, including expression on monocytes, dendritic cells, endothelial cells and epithelial cells. Therefore it is now thought that CD40 plays a more general role in immune regulation. The present paper reviews recent developments in this field of research with an emphasis on its expression and function on cells other than B lymphocytes (see Biotoon pp. 220-221), on CD40 signal transduction and on the function of CD40-CD40L in transplant rejection.

ABBREVIATIONS:

TRAF	TNF receptor associated factor
Jak3	Janus protein tyrosine kinases
ERK	p44 MAP kinase
p38	p38 MAP kinase
JNK	c-Jun NH2-terminal kinase
STAT	signal transducers and activators of transcription
NFκB	nuclear factor kappa B
NFAT	nuclear factor of activated T cells

Expression and Function of CD40 on Hematopoietic Cells

B Lymphocytes. Extensive studies on CD40 activation of B cells *in vitro*, have demonstrated that CD40 activation has major effects on many steps of the B cell natural history.¹⁻³ CD40 ligation activates resting B cells as shown by their increase in size and expression of new surface molecules involved in homotypic and heterotypic aggregation (CD23, VLA-4), T cell costimulation (CD80/CD86) and increased expression of MHC class I and II molecules and the TAP transporter. Furthermore, CD40-activated B cells secrete a myriad of cytokines which may act as autocrine and paracrine growth as well as differentiation factors. CD40 acts in concert with either cytokines or other receptor-ligand interactions for most biological functions of B cells, including the generation of germinal centers, isotype switching, induction of somatic mutations and selection of high affinity antibodies.

Monocytes and Dendritic Cells. Primary human monocytes either freshly isolated or cultured for 48 h show low but detectable CD40 surface protein expression.⁴ Strong expression is observed on cultured plastic adherent human monocytes.⁵ Monocytes stimulated by CD40L transfected cells secrete low

amounts of IL-6 and IL-8 and turn on their tumoricidal activity against a melanoma cell line.^{4,6} Finally, CD40 ligation activates NO synthesis as shown by the inhibition of T-cell-induced nitric oxide production by mouse macrophages using anti-CD40 antibodies.⁷

The expression of CD40 on antigen presenting cells like monocytes and dendritic cells has an important role *in vivo*.^{8,9} Activation of the CD40 receptors is one of the critical signals which allow the full maturation of dendritic cells into the most powerful antigen presenting cells.¹⁰ CD40 ligation on dendritic cells results in:

- increased expression of costimulatory molecules such as CD54/ICAM-1, CD58/LFA-3, CD80/B7-1, CD86/B7-2;
- the secretion of cytokines (such as IL1, IL6, IL8, IL10, IL12, TNF-α, MIP1α) and enzymes such as matrix metalloproteinase (MMP);
- an enhanced survival of these cells
- NO synthesis;
- promote differentiation of DC from CD34⁺ hematopoietic progenitors.¹¹⁻¹⁴

Therefore the interaction between CD40 and CD40L has important consequences for both APC function and T cell function. The full impact of

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CD40 for DC biology is outside the scope of this paper and is reviewed elsewhere.^{15,16}

T Lymphocytes. CD40 expression has been observed on rare CD4⁺ and CD8⁺ T cell clones,¹⁷ as well as on T cells isolated from patients with rheumatoid arthritis.¹⁸ Human resting T lymphocytes activated by a soluble trimeric CD40L or by CD40L transfected cells express activation molecules such as CD25, CD69 and CD40L.¹⁹ Both $\alpha\beta$ and $\gamma\delta$ mouse T cells are stimulated in a similar way, resulting in enhanced proliferation, induction of activation markers (CD25, CD69) and secretion of cytokines (IL-2, IFN- γ and TNF- α).^{20,21}

CD40 on Non-Hematopoietic Cells

Apart from hematopoietic cells, the expression of CD40 has now been observed on many other cell lineages. Although the expression is generally low in normal tissues, the molecule is clearly upregulated under various pathological conditions. In accordance, expression can be observed on many cultured cell types including endothelial cells, epithelial cells and fibroblasts.

Endothelial Cells. Endothelial cells of microvessels in the skin express CD40 intensely.^{22,23} In agreement, cultured human umbilical vein endothelial cells (HUVEC) constitutively express the CD40 antigen. The expression of CD40 is significantly upregulated by IL-1 α , TNF- α , IFN- γ and IFN- β , but not by IL-4.²³ Crosslinking of CD40 on cultured HUVEC results in a marked upregulation of ICAM-1 (CD54), VCAM-1 (CD106) and E-selectin (CD62E), thus resulting in an increased ability to bind leukocytes.^{22,23} Activation of endothelial cells also results in increased production of inflammatory mediators.^{24,25} Interaction between endothelial cells and T cells might also result in a positive feedback for CD40L expression via CD2/LFA3 expression.²⁶

Interestingly, CD40 expression on endothelial and other vascular wall cells such as smooth muscle cells is important for the development of atherosclerosis.²⁷ In vitro activation of these cells via CD40 results in expression of tissue factor,²⁸ interleukin-1 converting enzyme (ICE; caspase-1)²⁹ and metalloproteinases, including stromelysin-3.^{30,31} An important remaining question is the source of CD40L in these vascular lesions, which might include activated T cells, vascular cells themselves,³² or maybe more likely, activated platelets.³³

Epithelial Cells. The CD40 antigen was identified initially with the mAb S2C6 which was generated from mice immunized with a urinary bladder carcinoma.³⁴ Subsequently, the CD40 antigen has been identified on carcinomas of other origin such as colon, prostate, breast and lung, as well as on melanomas. Interestingly, on carcinomas it has been demonstrated that CD40 activation results in induction of cell death.³⁵⁻³⁷ Similarly CD40 ligation appears to inhibit the proliferation of diffuse B cell lymphomas.^{38,39} and could therefore provide a new therapeutic tool in some specific tumor cases.

CD40 is also expressed on non-malignant epithelial cells, including the CD45-negative stromal cell population in cortex and medulla of the human thymus.⁴⁰ IL-1 α , TNF- α and IFN- γ significantly upregulate CD40 levels on cultured epithelial cells, and these cells are induced to secrete GM-CSF upon CD40 ligation.⁴⁰ CD40 expression on thymic epithelial cells might also provide a costimulatory signal for human thymocytes.⁴¹

In renal interstitial inflammation, as during allograft rejection, tubular epithelial cells (TEC) play an active role in the production of pro-inflammatory mediators and a wide variety of chemokines.⁴² In normal tissue, only low expression of CD40 is observed on tubules, but it is strongly increased under inflammatory conditions. In vitro, tubular CD40 activation enhances the production of chemokines like IL-8, MCP-1 and RANTES.⁴³ More importantly, simultaneous addition of pro-inflammatory cytokines like IL-1 or IL-17 showed strong synergistic effects on the production of these mediators.^{44,45} Moreover, it was demonstrated that the cytokines IL-4 and IL-13 specifically enhanced CD40-induced RANTES production,⁴⁶ whereas they did not affect the production of other chemokines. These data suggest a complex network of interactions between epithelial cells and activated T cells, as also shown by in vitro cocultures.⁴⁷

Networks of interaction and crosstalk between activated T cells and epithelial cells via CD40-CD40L is most likely not restricted to the kidney. For instance, CD40 is also expressed on skin keratinocytes and crosslinking of the receptor results in further differentiation of these cells and an increased IL-6 production.⁴⁸⁻⁵⁰

Fibroblasts and Other Non-Hematopoietic Cells. Tissue fibrosis is characterized by the local presence of infiltrating mononuclear cells and activation of

DENDRITIC CELLS:

Antigen-presenting cells which play a critical role in the induction of immune responses, but are also instrumental for the induction of tolerance. Subsets of DC have been identified, but the exact relations between functions, lineage, origin and activation states are still a matter of debate.

TUBULAR EPITHELIAL CELLS:

Epithelial cells in the kidney which not only play a critical role in water and salt homeostasis, but are also an active participant in local inflammatory responses.

fibroblasts.⁵¹ CD40 is expressed on various fibroblast populations, including cells derived from skin, lung, kidney and synovium. Ligation of CD40 on synovial fibroblasts results in an enhanced growth, expression of adhesion molecules and production of various cytokines, including IL-6, GM-CSF and VEGF.⁵²⁻⁵⁴ Lung fibroblasts start to express higher levels of cyclooxygenase-2 upon CD40 activation.⁵⁵ It has been suggested that expression of CD40 on fibroblasts from different organs might associate with different chemokine expression profiles.⁵⁶ Therefore, cells of different origin should be studied separately, since they might have different biological potential.

CD40 is expressed on follicular dendritic cells (FDC) of secondary lymphoid organs as shown by immunohistology and also on freshly isolated FDC. Interaction between activated T cells and an FDC-like cell line results in enhanced growth and increased expression of ICAM-1, mediated via CD40-CD40L interaction.⁵⁷

Reed Sternberg (RS) cells, characteristic of Hodgkin's lymphoma, display enhanced clonogenic capacity and colony cell survival after engagement of CD40 by soluble CD40L.⁵⁸ Crosslinking CD40 results in enhanced production of IL-6, IL-8 TNF- α and LT α , as well as upregulation of ICAM-1 and CD80.⁵⁹

Finally, there are reports on CD40 expression on human muscle cells, regulating the production of various cytokines,⁶⁰ whereas crosslinking of CD40 expressed on human hepatocytes results in the induction of apoptosis via the Fas-FasL pathway.⁶¹ Understanding the different biological functions of CD40 on different cell types will depend on the insight in mechanisms of CD40 signal transduction.

CD40 Signal Transduction

Coupling of CD40 to different signaling pathways has been better understood by the identification of a family of associated proteins: TRAF (TNFR associated factor)⁶² At least four TRAF molecules have been demonstrated to associate with the CD40 cytoplasmic tail (TRAF2, TRAF3, TRAF5, TRAF6). Deletion mutants of the intracellular region of CD40 showed association of TRAF2, TRAF3 and TRAF5 to one region (residue 246-269), whereas TRAF6 associated with a separate domain (residue 230-245)⁶³ In accordance, pepscan analysis of CD40 identified PVQET (residue 250-254) as the minimal unit for TRAF1, TRAF2 and TRAF3

binding, and QEPQEINF (residue 231-238) as the minimum for TRAF6 binding (Fig. 1).^{64,65} To unravel this complex network and to understand the *in vivo* role of the different TRAF molecules, several members have been genetically inactivated in mice.

Genetic inactivation of TRAF3, TRAF2 and TRAF5 respectively, revealed no gross abnormalities at birth, but the mice died early in life.⁶⁶⁻⁶⁸ Functional analysis of signaling pathways showed alterations in NF- κ B and JNK activation and the expression of several activation markers, but suggested a certain level of redundancy. Generation of TRAF6 KO mice showed low birth rates of *-/-* mice, severe osteopetrosis and early death.^{69,70} TRAF6 is also critical in the signal transduction induced by the Toll/IL1-R family.⁷¹⁻⁷⁴ Accordingly, the absence of TRAF6 not only interferes with CD40-mediated B cell proliferation, but also B cell proliferation induced by LPS or induction of iNOS in macrophages by IL-1 or LPS.^{69,70}

Recent studies identified a domain (pre-ligand assembly domain; PLAD) in the extracellular part of CD40, which is responsible for the assembly of a trimeric receptor complex in the absence of a ligand.⁷⁵ It remains to be determined whether there is a signaling function for ligand-independent trimers and what is the exact consequence of ligand binding. Upon activation, CD40 together with associated TRAF molecules move to lipid microdomains which brings them in close vicinity of different src kinases.^{76,77} A wide variety of kinases have been implicated to become activated by the CD40 pathway, including PTKs, PKA, PI3K, JAK3, NIK, JNK, ERK and p38 MAPK. However, it remains to be established which pathways are important under certain conditions or in specific cell types. For instance, although the NF- κ B inducing kinase (NIK) was shown to be essential for B-cell activation, it seems to be dispensable for activation of dendritic cells.⁷⁸ Similarly, TRAF6 seems to be an intermediate in CD40-induced NF- κ B activation in 293 epithelial cells, but not in B cells.⁷⁹

CD40 and Transplantation

Interference with CD40 activation prolongs the survival of experimentally transplanted organs. These models have been mostly limited to murine models, including heart, skin, aorta and pancreatic islets allografts, as well as the respective xenografts.⁸⁰⁻⁸³ The contribution of CD40-

TRAF:

A group of intracellular proteins, TNF-receptor associated factors, which on the one hand associate with the intracellular parts of receptors of the TNFR family, including CD40, and on the other hand connect to different signal transduction pathways.

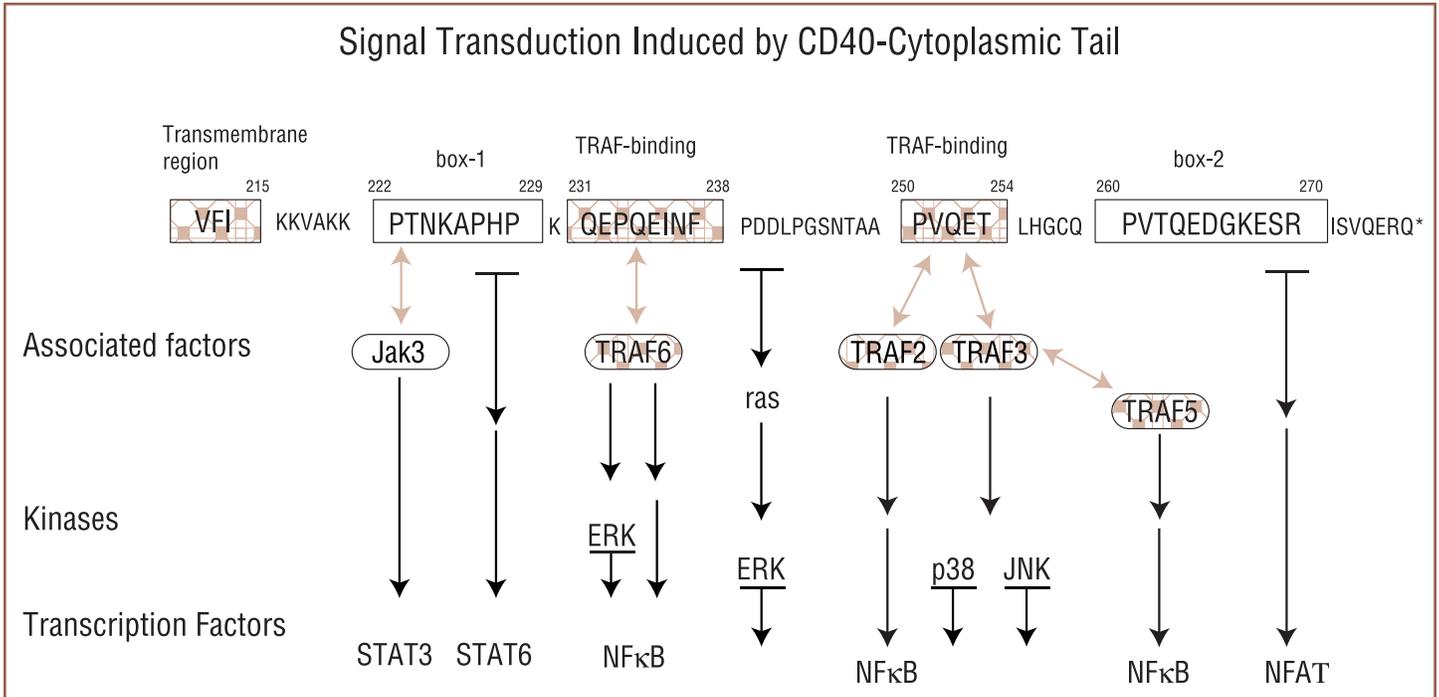


Figure 1. Schematic representation of the amino acid sequence of the intracellular region of human CD40 (single letter code and numbering of mature protein). Several domains have been identified which either directly associate with identified adaptor molecules such as TRAFs or which have been implicated in the induction of specific pathways, ultimately leading to gene activation.

CHEMOKINES:

A group of small polypeptides which are responsible for the chemoattraction of leukocytes to sites of inflammation, or in the normal trafficking of these cells. Specific interaction with chemokine receptors, which are differentially expressed on different cell types, will determine the composition of cellular infiltrate.

HUMANIZED ANTI-CD40L:

A monoclonal antibody specific for CD40L, which was initially raised in mice, but with all constant regions of the Ig molecule replaced for the human sequence. Only the antigen recognition site is still of mouse origin which should diminish the in vivo anti-mouse response observed with classical mouse monoclonal antibodies.

CD40L interactions in the process of T cell priming, differentiation and effector functions has been extensively reviewed.^{9,84-86} However, it has been shown that the results in transplantation models are strongly dependent on the strain combinations used. In several cases, rejection seems to occur independent of CD40-CD40L and is mediated by CD8 positive T cells.^{87,88} Moreover, it has been shown that the Th1 or Th2 background of the mice affects the result of anti-CD40L blockade.⁸⁹ Treatment regimens have been different in the various studies, ranging from once at the time of transplantation to repeated administration every 72 hours to prevent chronic rejection in aortic allografts.⁹⁰ Local expression of CD40L has been found in human renal and heart allografts, and its expression seems to correlate with rejection.^{45,91-93} Studies with a humanized anti-CD40L antibody have been performed in renal allotransplantation in rhesus monkeys. An extended course of anti-CD40L as the only treatment over a 6 months period resulted in long-term survival.^{94,95} Importantly, the organs were not rejected when treatment was stopped, and graft survival in this

group reached up to 10 months post-treatment. Combinations of current immunosuppressive drugs, including steroids, MMF, and FK506, seem to counteract the beneficial effect of anti-CD40L. The same humanized Ab has been used in a primate model of cardiac transplantation, showing improved survival and less vascular pathology.⁹⁶

Conclusion

Research on CD40 has over the years shifted from a restricted role in the regulation of humoral immunity to a much broader role in immune activation and inflammation. Although the present data do not provide sufficient evidence to show permanent transplant tolerance, the data are promising and have resulted in a phase II human clinical trial in renal allograft recipients. Despite discontinuation of this trial due to thrombo-embolic complications,⁹⁷ it is expected to be resumed. In view of the broad expression of the CD40 receptor on many cells with a potential role in allograft rejection, further detailed analysis is needed to unravel the mechanism of action of CD40L blockade.

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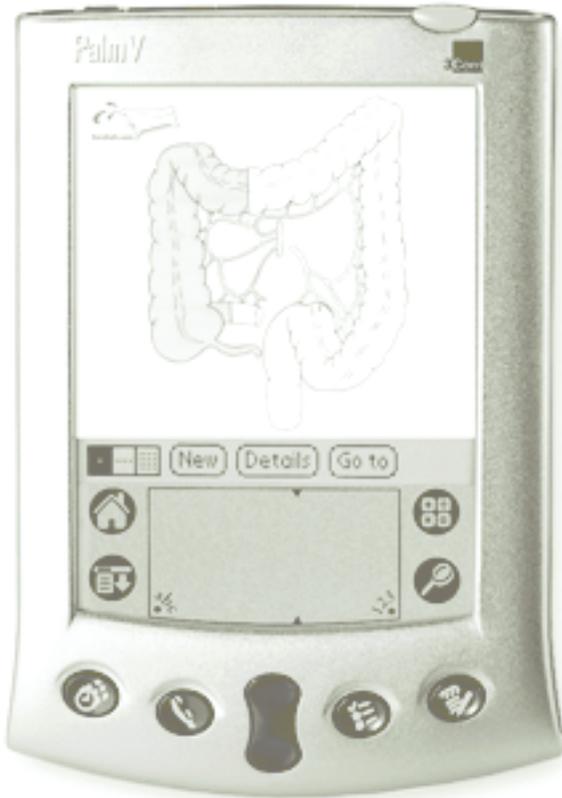
REFERENCES

- Banchereau J, Bazan F, Blanchard D et al. The CD40 antigen and its ligand. *Annu Rev Immunol* 1994; **12**:881-922.
- Van Kooten C, Banchereau J. CD40-CD40 ligand: A multifunctional receptor-ligand pair. *Adv Immunol* 1996; **61**:1-77.
- Foy TM, Aruffo A, Bajorath J et al. Immune regulation by CD40 and its ligand gp39. *Annu Rev Immunol* 1996; **14**:591-617.
- Alderson MR, Armitage RJ, Tough TW et al. CD40 expression by human monocytes: Regulation by cytokines and activation of monocytes by the ligand for CD40. *J Exp Med* 1993; **178**:669-74.
- Wagner DJ, Stout RD, Suttles J. Role of the CD40-CD40 ligand interaction in CD4⁺ T cell contact-dependent activation of monocyte interleukin-1 synthesis. *Eur J Immunol* 1994; **24**:3148-54.
- Kiener PA, Moran-Davis P, Rankin BM et al. Stimulation of CD40 with purified soluble gp39 induces proinflammatory responses in human monocytes. *J Immunol* 1995; **155**:4917-25.
- Tian L, Noelle RJ, Lawrence DA. Activated T cells enhance nitric oxide production by murine splenic macrophages through gp39 and LFA-1. *Eur J Immunol* 1995; **25**:306-9.
- Stout RD, Suttles J. The many roles of CD40 in cell-mediated inflammatory responses. *Immunol Today* 1996; **17**:487-92.
- Grewal IS, Flavell RA. A central role of CD40 ligand in the regulation of CD4⁺ T cell responses. *Immunol Today* 1996; **17**:410-4.
- Banchereau J, Steinman RM. Dendritic cells and the control of immunity. *Nature* 1998; **392**:245-52.
- Caux C, Massacrier C, Vanbervliet B et al. Activation of human dendritic cells through CD40 crosslinking. *J Exp Med* 1994; **180**:1263-72.
- Cella M, Scheidegger D, Palmer-Lehmann K et al. Ligation of CD40 on dendritic cells triggers production of high levels of interleukin-12 and enhances T-cell stimulatory capacity: T-T help via APC activation. *J Exp Med* 1996; **184**:747-52.
- Koch F, Stanzl U, Jennewein P et al. High level IL-12 production by murine dendritic cells: upregulation via MHC class II and CD40 molecules and downregulation by IL-4 and IL-10. *J Exp Med* 1996; **184**:741-6.
- Flores-Romo L, Bjorck P, Duvert V et al. CD40 ligation on human cord blood CD34⁺ hematopoietic progenitors induces their proliferation and differentiation into functional dendritic cells. *J Exp Med* 1997; **185**:341-9.
- Lane PJ, Brocker T. Developmental regulation of dendritic cell function. *Curr Opin Immunol* 1999; **11**:308-13.
- Banchereau J, Briere F, Caux C et al. Immunobiology of dendritic cells. *Annu Rev Immunol* 2000; **18**:767-811.
- Ware CF, Crowe PD, Vanarsdale TL et al. Tumor necrosis factor (TNF) receptor expression in T lymphocytes. Differential regulation of the Type I TNF receptor during activation of resting and effector T cells. *J Immunol* 1991; **147**:4229-38.
- Potocnik AJ, Kinne R, Menninger H et al. Expression of activation antigens on T cells in rheumatoid arthritis patients. *Scand J Immunol* 1990; **31**:213-24.
- Armitage RJ, Tough TW, Macduff BM et al. CD40 ligand is a T cell growth factor. *Eur J Immunol* 1993; **23**:2326-31.
- Fanslow WC, Clifford KN, Seaman M et al. Recombinant CD40 ligand exerts potent biologic effects on T cells. *J Immunol* 1994; **152**:4262-9.
- Ramsdell F, Seaman MS, Clifford KN et al. CD40 ligand acts as a costimulatory signal for neonatal thymic gamma delta T cells. *J Immunol* 1994; **152**:2190-7.
- Hollenbaugh D, Mischel-Petty N, Edwards CP et al. Expression of functional CD40 by vascular endothelial cells. *J Exp Med* 1995; **182**:33-40.
- Karmann K, Hughes CC, Schechner J et al. CD40 on human endothelial cells: Inducibility by cytokines and functional regulation of adhesion molecule expression. *Proc Natl Acad Sci USA* 1995; **92**:4342-6.
- Yellin MJ, Brett J, Baum D et al. Functional interactions of T cells with endothelial cells: The role of CD40L-CD40-mediated signals. *J Exp Med* 1995; **182**:1857-64.
- Dechanet J, Grosset C, Taupin JL et al. CD40 ligand stimulates proinflammatory cytokine production by human endothelial cells. *J Immunol* 1997; **159**:5640-7.
- Karmann K, Hughes CC, Fanslow WC et al. Endothelial cells augment the expression of CD40 ligand on newly activated human CD4⁺ T cells through a CD2/LFA-3 signaling pathway. *Eur J Immunol* 1996; **26**:610-7.
- Mach F, Schonbeck U, Sukhova GK et al. Reduction of atherosclerosis in mice by inhibition of CD40 signalling. *Nature* 1998; **394**:200-3.
- Schonbeck U, Mach F, Sukhova GK et al. CD40 ligation induces tissue factor expression in human vascular smooth muscle cells. *Am J Pathol* 2000; **156**:7-14.
- Schonbeck U, Mach F, Bonnefoy JY et al. Ligation of CD40 activates Interleukin 1 β -converting enzyme (caspase-1) activity in vascular smooth muscle and endothelial cells and promotes elaboration of active Interleukin 1 β . *J Biol Chem* 1997; **272**:19569-74.
- Schonbeck U, Mach F, Sukhova GK et al. Expression of stromelysin-3 in atherosclerotic lesions: Regulation via CD40-CD40 ligand signaling. *In vitro and In vivo*. *J Exp Med* 1999; **189**:843-53.
- Mach F, Schonbeck U, Fabunmi RP et al. T lymphocytes induce endothelial cell matrix metalloproteinase expression by a CD40L-dependent mechanism: Implications for tubule formation. *Am J Pathol* 1999; **154**:229-38.
- Mach F, Schonbeck U, Sukhova GK et al. Functional CD40 ligand is expressed on human vascular endothelial cells, smooth muscle cells, and macrophages: implications for CD40-CD40 ligand signaling in atherosclerosis. *Proc Natl Acad Sci USA* 1997; **94**:1931-6.
- Henn V, Slupsky JR, Grafe M et al. CD40 ligand on activated platelets triggers an inflammatory reaction of endothelial cells. *Nature* 1998; **391**:591-4.
- Koho H, Paulie S, Ben-Aissa H et al. Monoclonal antibodies to antigens associated with transitional cell carcinoma of the human urinary bladder. I. Determination of the selectivity of six antibodies by cell ELISA and immunofluorescence. *Cancer Immunol Immunother* 1984; **17**:165-72.
- Hess S, Engelmann H. A novel function of CD40: Induction of cell death in transformed cells. *J Exp Med* 1996; **183**:159-67.
- Young LS, Eliopoulos AG, Gallagher NJ et al. CD40 and epithelial cells: Across the great divide. *Immunol Today* 1998; **19**:502-6.
- Hirano A, Longo DL, Taub DD et al. Inhibition of human breast carcinoma growth by a soluble recombinant human CD40 ligand. *Blood* 1999; **93**:2999-3007.
- Murphy WJ, Funakoshi S, Beckwith M et al. Antibodies to CD40 prevent Epstein-Barr virus-mediated human B cell lymphomagenesis in severe combined immune deficient mice given human peripheral blood lymphocytes. *Blood* 1995; **86**:1946-53.
- Costello RT, Gastaut JA, Olive D. What is the real role of CD40 in cancer immunotherapy? *Immunol Today* 1999; **20**:488-93.
- Galy AHM, Spits H. CD40 is functionally expressed on human thymic epithelial cells. *J Immunol* 1992; **149**:775-82.
- Ruggiero G, Caceres EM, Voordouw A et al. CD40 expressed on thymic epithelial cells provides costimulation for proliferation but not for apoptosis of human thymocytes. *J Immunol* 1996; **156**:3737-46.
- Van Kooten C, Daha MR, Van Es LA. Tubular epithelial cells: A critical cell type in the regulation of renal inflammatory processes. *Exp Nephrol* 1999; **7**:429-37.
- Van Kooten C, Gerritsma JSJ, Paape ME et al. Possible role for CD40-CD40L in the regulation of interstitial infiltration in the kidney. *Kidney Int* 1997; **51**:711-21.

44. Van Kooten C, Van der Linde X, Woltman AM et al. Synergistic effect of IL-1 and CD40L on the activation of human renal tubular epithelial cells. **Kidney Int** 1999; **56:41-51**.
45. Woltman AM, De Haij S, Boonstra JG et al. Interleukin-17 and CD40-ligand synergistically enhance cytokine and chemokine production by renal epithelial cells. **J Am Soc Nephrol** 2000; **11:2044-55**.
46. Deckers JGM, De Haij S, Van der Woude FJ et al. IL-4 and IL-13 augment cytokine- and CD40-induced RANTES production by human renal tubular epithelial cells in vitro. **J Am Soc Nephrol** 1998; **9:1187-93**.
47. Kuroiwa T, Schlimgen R, Illei GG et al. Distinct T cell/renal tubular epithelial cell interactions define differential chemokine production: Implications for tubulointerstitial injury in chronic glomerulonephritides. **J Immunol** 2000; **164:3323-9**.
48. Gaspari AA, Sempowski GD, Chess P et al. Human epidermal keratinocytes are induced to secrete interleukin-6 and co-stimulate T lymphocyte proliferation by a CD40-dependent mechanism. **Eur J Immunol** 1996; **26:1371-7**.
49. Denfeld RW, Hollenbaugh D, Fehrenbach A et al. CD40 is functionally expressed on human keratinocytes. **Eur J Immunol** 1996; **26:2329-34**.
50. Peguet-Navarro J, Dalbiez-Gauthier C, Moulon C et al. CD40 ligation of human keratinocytes inhibits their proliferation and induces their differentiation. **J Immunol** 1997; **158:144-52**.
51. Phipps RP. Atherosclerosis: The emerging role of inflammation and the CD40-CD40 ligand system. **Proc Natl Acad Sci USA** 2000; **97:6930-2**.
52. Yellin MJ, Winikoff S, Fortune SM et al. Ligation of CD40 on fibroblasts induces CD54 (ICAM-1) and CD106 (VCAM-1) upregulation and IL-6 production and proliferation. **J Leuk Biol** 1995; **58:209-16**.
53. Rissoan MC, Van Kooten C, Chomarat P et al. The functional CD40 antigen of fibroblasts may contribute to the proliferation of rheumatoid synovium. **Clin Exp Immunol** 1996; **106:481-90**.
54. Cho CS, Cho ML, Min SY et al. CD40 engagement on synovial fibroblast upregulates production of vascular endothelial growth factor. **J Immunol** 2000; **164:5055-61**.
55. Zhang Y, Cao HJ, Graf B et al. CD40 engagement upregulates cyclooxygenase-2 expression and prostaglandin E2 production in human lung fibroblasts. **J Immunol** 1998; **160:1053-7**.
56. Brouty-Boye D, Pottin-Clemenceau C, Doucet C et al. Chemokines and CD40 expression in human fibroblasts. **Eur J Immunol** 2000; **30:914-9**.
57. Kim HS, Zhang X, Choi YS. Activation and proliferation of follicular dendritic cell-like cells by activated T lymphocytes. **J Immunol** 1994; **153:2951-61**.
58. Carbone A, Gloghini A, Gattei V et al. Expression of functional CD40 antigen on Reed-Sternberg cells and Hodgkin's disease cell lines. **Blood** 1995; **85:780-9**.
59. Gruss HJ, Hirschstein D, Wright B et al. Expression and function of CD40 on Hodgkin and Reed-Sternberg cells and the possible relevance for Hodgkin's disease. **Blood** 1994; **84:2305-14**.
60. Sugiura T, Kawaguchi Y, Harigai M et al. Increased CD40 expression on muscle cells of polymyositis and dermatomyositis: role of CD40-CD40 ligand interaction in IL-6, IL-8, IL-15, and monocyte chemoattractant protein-1 production. **J Immunol** 2000; **164:6593-600**.
61. Afford SC, Randhawa S, Eliopoulos AG et al. CD40 activation induces apoptosis in cultured human hepatocytes via induction of cell surface fas ligand expression and amplifies fas-mediated hepatocyte death during allograft rejection [In Process Citation]. **J Exp Med** 1999; **189:441-6**.
62. Inoue J, Ishida T, Tsukamoto N et al. Tumor necrosis factor receptor-associated factor (TRAF) family: adapter proteins that mediate cytokine signaling. **Exp Cell Res** 2000; **254:14-24**.
63. Ishida T, Mizushima S, Azuma S et al. Identification of TRAF6, a novel tumor necrosis factor receptor-associated factor protein that mediates signaling from an amino-terminal domain of the CD40 cytoplasmic region. **J Biol Chem** 1996; **271:28745-8**.
64. Pullen SS, Miller HG, Everdeen DS et al. CD40-tumor necrosis factor receptor-associated factor (TRAF) interactions: regulation of CD40 signaling through multiple TRAF binding sites and TRAF hetero-oligomerization. **Biochemistry** 1998; **37:11836-45**.
65. Leo E, Welsh K, Matsuzawa S et al. Differential requirements for tumor necrosis factor receptor-associated factor family proteins in CD40-mediated induction of NF-kappaB and Jun N-terminal kinase activation. **J Biol Chem** 1999; **274:22414-22**.
66. Xu Y, Cheng G, Baltimore D. Targeted disruption of TRAF3 leads to post-natal lethality and defective T-dependent immune responses. **Immunity** 1996; **5:407-15**.
67. Yeh WC, Shahinian A, Speiser D et al. Early lethality, functional NF-kappaB activation, and increased sensitivity to TNF-induced cell death in TRAF2-deficient mice. **Immunity** 1997; **7:715-25**.
68. Nakano H, Sakon S, Koseki H et al. Targeted disruption of Traf5 gene causes defects in CD40- and CD27-mediated lymphocyte activation. **Proc Natl Acad Sci USA** 1999; **96:9803-8**.
69. Lomaga MA, Yeh WC, Sarosi I et al. TRAF6 deficiency results in osteopetrosis and defective interleukin-1, CD40, and LPS signaling. **Genes Dev** 1999; **13:1015-24**.
70. Naito A, Azuma S, Tanaka S et al. Severe osteopetrosis, defective interleukin-1 signalling and lymph node organogenesis in TRAF6-deficient mice. **Genes Cells** 1999; **4:353-62**.
71. Cao Z, Xiong J, Takeuchi M et al. TRAF6 is a signal transducer for interleukin-1. **Nature** 1996; **383:443-6**.
72. Rock FL, Hardiman G, Timans JC et al. A family of human receptors structurally related to *Drosophila* Toll. **Proc Natl Acad Sci USA** 1998; **95:588-93**.
73. Adachi O, Kawai T, Takeda K et al. Targeted disruption of the MyD88 gene results in loss of IL-1- and IL-18-mediated function. **Immunity** 1998; **9:143-50**.
74. Kawai T, Adachi O, Ogawa T et al. Unresponsiveness of MyD88-deficient mice to endotoxin. **Immunity** 1999; **11:115-22**.
75. Chan FK, Chun HJ, Zheng L et al. A domain in TNF receptors that mediates ligand-independent receptor assembly and signaling. **Science** 2000; **288:2351-4**.
76. Hostager BS, Catlett IM, Bishop GA. Recruitment of CD40 and tumor necrosis factor receptor-associated factors 2 and 3 to membrane microdomains during CD40 signaling. **J Biol Chem** 2000; **275:15392-8**.
77. Vidalain PO, Azocar O, Servet-Delprat C et al. CD40 signaling in human dendritic cells is initiated within membrane rafts. **Embo J** 2000; **19:3304-13**.
78. Garceau N, Kosaka Y, Masters S et al. Lineage-restricted function of nuclear factor kappaB-inducing kinase (NIK) in transducing signals via CD40. **J Exp Med** 2000; **191:381-6**.
79. Jalukar SV, Hostager BS, Bishop GA. Characterization of the roles of TNF receptor-associated factor 6 in CD40-mediated B lymphocyte effector functions. **J Immunol** 2000; **164:623-30**.
80. Larsen CP, Pearson TC. The CD40 pathway in allograft rejection, acceptance, and tolerance. **Curr Opin Immunol** 1997; **9:641-7**.
81. Sayegh MH, Turka LA. The role of T-cell costimulatory activation pathways in transplant rejection. **N Engl J Med** 1998; **338:1813-21**.
82. Gudmundsdottir H, Turka LA. T cell costimulatory blockade: New therapies for transplant rejection. **J Am Soc Nephrol** 1999; **10:1356-65**.
83. Van Kooten C. Blockade of costimulatory pathways of T cell activation: The solution to acute and chronic rejection? **Curr Opin Nephrol Hypertens** 1999; **8:675-80**.
84. Stout RD, Suttles J, Xu J et al. Impaired T cell-mediated macrophage activation in CD40 ligand-deficient mice. **J Immunol** 1996; **156:8-11**.
85. Mackey MF, Barth RJ, Jr., Noelle RJ. The role of CD40/CD154 interactions in the priming, differentiation, and effector function of helper and cytotoxic T cells. **J Leukoc Biol** 1998; **63:418-28**.
86. Heath WR, Carbone FR. Cytotoxic T lymphocyte activation by crosspriming. **Curr Opin Immunol** 1999; **11:314-8**.
87. Trambley J, Bingaman AW, Lin A et al. Asialo GM1+ CD8+ T cells play a critical role in costimulation blockade-resistant allograft rejection. **J Clin Invest** 1999; **104:1715-22**.
88. Jones ND, Van Maurik A, Hara M et al. CD40-CD40 ligand-independent activation of CD8+ T cells can trigger allograft rejection. **J Immunol** 2000; **165:1111-8**.

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89. Kishimoto K, Dong VM, Issazadeh S et al. The role of CD154-CD40 versus CD28-B7 costimulatory pathways in regulating allogeneic Th1 and Th2 responses in vivo. **J Clin Invest** 2000; **106:63-72**.
90. Sun H, Subbotin V, Chen C et al. Prevention of chronic rejection in mouse aortic allografts by combined treatment with CTLA4-Ig and anti-CD40 ligand monoclonal antibody. **Transplantation** 1997; **64:1838-43**.
91. Reul RM, Fang JC, Denton MD et al. CD40 and CD40 ligand (CD154) are coexpressed on microvessels in vivo in human cardiac allograft rejection. **Transplantation** 1997; **64:1765-74**.
92. Zheng XX, Schachter AD, Vasconcellos L et al. Increased CD40 ligand gene expression during human renal and murine islet allograft rejection. **Transplantation** 1998; **65:1512-5**.
93. Gaweco AS, Mitchell BL, Lucas BA et al. CD40 expression on graft infiltrates and parenchymal CD154 (CD40L) induction in human chronic renal allograft rejection. **Kidney Int** 1999; **55:1543-52**.
94. Kirk AD, Harlan DM, Armstrong NN et al. CTLA4-Ig and anti-CD40 ligand prevent renal allograft rejection in primates. **Proc Natl Acad Sci USA** 1997; **94:8789-94**.
95. Kirk AD, Burkly LC, Batty DS et al. Treatment with humanized monoclonal antibody against CD154 prevents acute renal allograft rejection in nonhuman primates. **Nat Med** 1999; **5:686-93**.
96. Pierson RNr, Chang AC, Blum MG et al. Prolongation of primate cardiac allograft survival by treatment with Anti-CD40 ligand (CD154) antibody. **Transplantation** 1999; **68:1800-5**.
97. Kawai T, Andrews D, Colvin RB et al. Thromboembolic complications after treatment with monoclonal antibody against CD40 ligand. **Nat Med** 2000; **6:114**.